

## HYPEREXCITABILITY OF BLADDER AFFERENT NEURONS ASSOCIATED WITH REDUCED EXPRESSION OF POTASSIUM CHANNEL KV1.4 $\alpha$ -SUBUNIT IN RATS WITH SPINAL CORD INJURY

### Hypothesis / aims of study

Hyperexcitability of C-fibre bladder afferent pathways has been proposed as an important pathophysiological basis of neurogenic detrusor overactivity with spinal cord injury (SCI). However, the functional and molecular mechanisms inducing hyperexcitability of C-fibre bladder afferent neurons after SCI are not fully elucidated. We therefore examined changes in electrophysiological properties of bladder afferent neurons obtained from spinal transected rats, especially focusing on voltage-gated potassium ( $K^+$ ) channels and the expression levels of  $\alpha$ -subunits, which can form A-type  $K^+$  ( $K_A$ ) channels. Activation of  $K_A$  channels is known to reduce excitability of capsaicin-sensitive C-fibre bladder afferent neurons from spinal intact rats [1].

### Study design, materials and methods

Freshly dissociated L6-S1 dorsal root ganglia (DRG) neurons were prepared from female spinal intact and spinal transected (4 weeks after T9-T10 transection) SD rats. Whole cell patch-clamp recordings were performed on individual bladder afferent neurons, which were labelled by retrograde axonal transport of a fluorescent dye, Fast Blue (FB) injected into the bladder wall 7 days prior the dissociation. Since the majority (80%) of C-fibre bladder afferent neurons are known to be sensitive to capsaicin, capsaicin-sensitive neurons were selected for evaluation. The expression levels of Kv1.2 and 1.4  $\alpha$ -subunits were evaluated using immunohistochemical methods.

### Results

Capsaicin-sensitive bladder afferent neurons from spinal transected rats exhibited increased cell excitability evidenced by lower thresholds for spike activation ( $-26.4 \pm 1.3$  mV) and the increased number of action potentials ( $4.7 \pm 0.7$  spikes) during a 800 msec depolarizing pulse compare to spinal intact rats ( $-21.8 \pm 0.9$  mV and  $1.3 \pm 0.1$  spikes, respectively) (Fig. 1). The peak density of slowly-inactivating  $K_A$  currents during membrane depolarizations to 0 mV in capsaicin-sensitive bladder afferent neurons of spinal transected rats was significantly smaller ( $38.1 \pm 4.6$  pA/pF) than that from spinal intact rats ( $68.6 \pm 6.3$  pA/pF) (Fig. 2), and the inactivation curve of the  $K_A$  current was displaced to more hyperpolarized levels by  $\sim 10$  mV after spinal transection (Fig. 3). On the other hand, the sustained delayed-rectifier  $K^+$  current density was not altered after spinal transection (Fig. 2). The expression of Kv1.4  $\alpha$ -subunits, which can form  $K_A$  channels, was reduced in bladder afferent neurons from spinal transected rats compared to spinal intact rats (Fig. 4).

### Interpretation of results

These results indicate that: (1) capsaicin-sensitive bladder afferent neurons in spinal transected rats show hyperexcitability as evidenced by lower thresholds for spike activation and tonic firing pattern, (2) the density of slowly-inactivating  $K_A$  currents was reduced and the inactivation curve of  $K_A$  currents for spinal transected rats was displaced to more hyperpolarized levels in comparison with spinal intact rats, (3) the expression of Kv1.4  $\alpha$ -subunits in bladder afferent neurons was decreased after spinal transection. Thus, it is assumed that the excitability of capsaicin-sensitive C-fiber bladder afferent neurons is increased in association with reductions in  $K_A$  current size and Kv1.4  $\alpha$ -subunit expression in spinal transected rats.

### Concluding message

The reduction in  $K_A$  channel activity due to downregulation of the Kv1.4  $\alpha$ -subunit could contribute to hyperexcitability of C-fibre bladder afferent pathways, leading to detrusor overactivity in SCI. Thus, the Kv1.4  $\alpha$ -subunit could be a potential molecular target for treating OAB due to neurogenic detrusor overactivity.

Fig.1

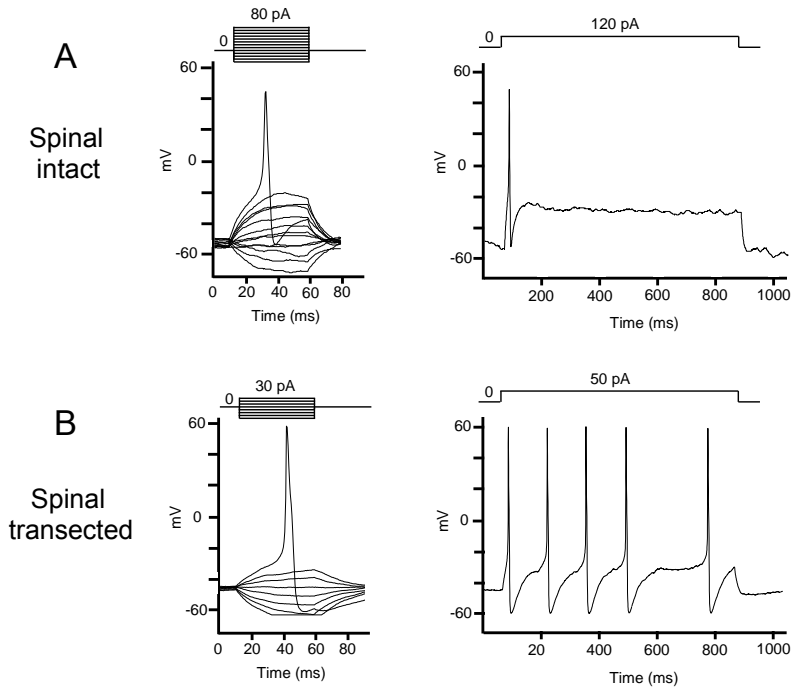


Fig.2

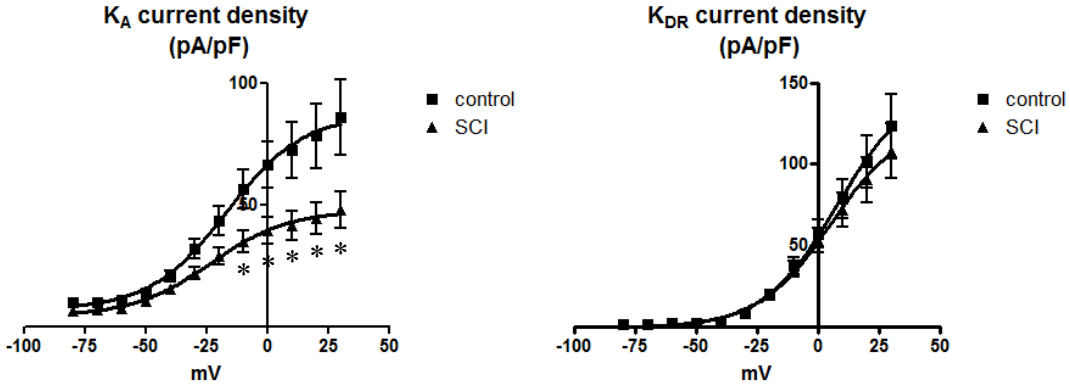
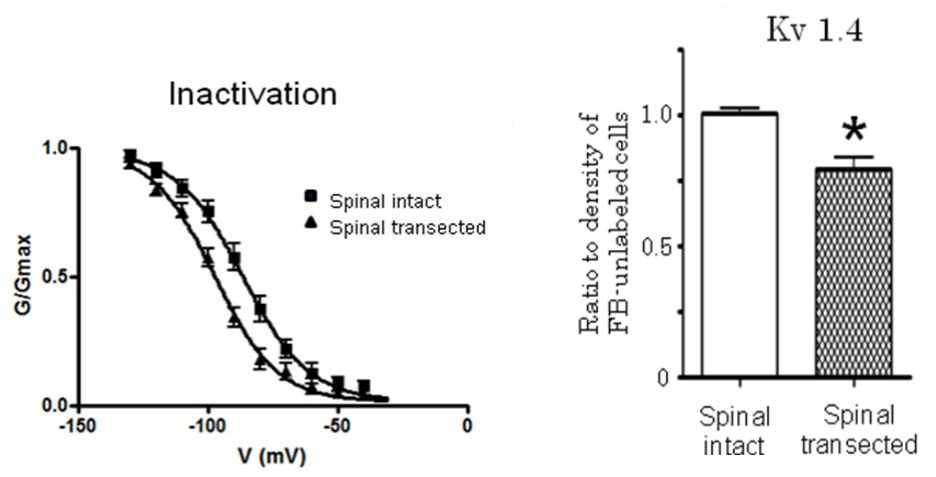


Fig.3

Fig.4



References

1. Yoshimura and de Groat, Journal of Neuroscience, 19: 4644-4653, 1999

Disclosures

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